

Influence of the Nitrogen Donors on Nonheme Iron Models of Superoxide Reductase: High-Spin Fe^{III}–OOR Complexes

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Abstract: A new five-coordinate, (N₄S(thiolate))Fe^{II} complex, containing tertiary amine donors, [Fe^{II}(Me₄[15]aneN₄)(SPh)]BPh₄ (**2**), was synthesized and structurally characterized as a model of the reduced active site of superoxide reductase (SOR). Reaction of **2** with *tert*-butyl hydroperoxide (*t*BuOOH) at –78 °C led to the generation of the alkylperoxo-iron(III) complex [Fe^{III}(Me₄[15]aneN₄)(SPh)(OO*t*Bu)]⁺ (**2a**). The nonthiolate-ligated complex, [Fe^{II}(Me₄[15]aneN₄)(OTf)₂] (**3**), was also reacted with *t*BuOOH and yielded the corresponding alkylperoxo complex [Fe^{III}(Me₄[15]aneN₄)(OTf)(OO*t*Bu)]⁺ (**3a**) at an elevated temperature of –23 °C. These species were characterized by low-temperature UV–vis, EPR, and resonance Raman spectroscopies. Complexes **2a** and **3a** exhibit distinctly different spectroscopic signatures than the analogous alkylperoxo complexes [Fe^{III}([15]aneN₄)(SAR)(OOR)]⁺, which contain secondary amine donors. Importantly, alkylation at nitrogen leads to a change from low-spin (*S* = 1/2) to high-spin (*S* = 5/2) of the iron(III) center. The resonance Raman data reveal that this change in spin state has a large effect on the $\nu(\text{Fe–O})$ and $\nu(\text{O–O})$ vibrations, and a comparison between **2a** and the nonthiolate-ligated complex **3a** shows that axial ligation has an additional significant impact on these vibrations. To our knowledge this study is the first in which the influence of a ligand trans to a peroxo moiety has been evaluated for a structurally equivalent pair of high-spin/low-spin peroxo-iron(III) complexes. The implications of spin state and thiolate ligation are discussed with regard to the functioning of SOR.

Introduction

The processing of dioxygen and related molecules, such as hydrogen peroxide and superoxide, is of critical importance to a number of heme and nonheme iron metalloproteins. In most of these systems the formation of iron–oxygen adducts in the form of iron-superoxo, -hydroperoxo, or -alkylperoxo species is a key step in their function. One such metalloprotein is superoxide reductase (SOR), which is found in anaerobic and microaerophilic organisms and reduces superoxide to hydrogen peroxide as part of the organism's natural defenses.^{1–10} The reduced, active state of SOR contains an Fe^{II} center bound by

four imidazolyl nitrogen donors from histidine arranged in a plane and a thiolato sulfur donor from cysteine in an axial position, resulting in a square pyramidal [(N₄S(thiolate))] geometry around the iron center. The coordination environment of SOR has an obvious structural similarity with the heme-iron active site of cytochrome P450, in which the iron center is coordinated by four pyrrolyl nitrogen donors in the equatorial plane and a cysteinate ligand in the axial position.^{11–14} The two enzymes are thought to access similar iron-(hydro)peroxy intermediates during catalysis, which then experience dramatically different fates; in the case of SOR the (hydro)peroxy ligand is released as H₂O₂, whereas in P450 O–O bond cleavage takes place and results in the formation of a reactive, high-valent iron-oxo species.

One possible factor that influences the fate of the iron(III)-(hydro)peroxo intermediate is the spin state of the iron center. Solomon and Que have performed spectroscopic studies, supported by theoretical calculations, on both hydroperoxo- and alkylperoxo-iron(III) model complexes that show that a low-

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- (1) Nivière, V.; Fontecave, M. *J. Biol. Inorg. Chem.* **2004**, *9*, 119–123.
- (2) Kurtz, D. M., Jr. *J. Inorg. Biochem.* **2006**, *100*, 679–693.
- (3) Dey, A.; Jenney, F. E.; Adams, M. W. W.; Johnson, M. K.; Hodgson, K. O.; Hedman, B.; Solomon, E. I. *J. Am. Chem. Soc.* **2007**, *129*, 12418–12431.
- (4) Huang, V. W.; Emerson, J. P.; Kurtz, D. M., Jr. *Biochemistry* **2007**, *46*, 11342–11351.
- (5) Kovacs, J. A.; Brines, L. M. *Acc. Chem. Res.* **2007**, *40*, 501–509.
- (6) Mathé, C.; Weill, C. O.; Mattioli, T. A.; Berthomieu, C.; Houée-Levin, C.; Tremey, E.; Nivière, V. *J. Biol. Chem.* **2007**, *282*, 22207–22216.
- (7) Moura, I.; Pauleta, S. R.; Moura, J. J. G. *J. Biol. Inorg. Chem.* **2008**, *13*, 1185–1195.
- (8) Katona, G.; Carpentier, P.; Nivière, V.; Amara, P.; Adam, V.; Ohana, J.; Tsanov, N.; Bourgeois, D. *Science* **2007**, *316*, 449–453.
- (9) Clay, M. D.; Yang, T. C.; Jenney, F. E.; Kung, I. Y.; Cosper, C. A.; Krishnan, R.; Kurtz, D. M., Jr.; Adams, M. W. W.; Hoffman, B. M.; Johnson, M. K. *Biochemistry* **2006**, *45*, 427–438.

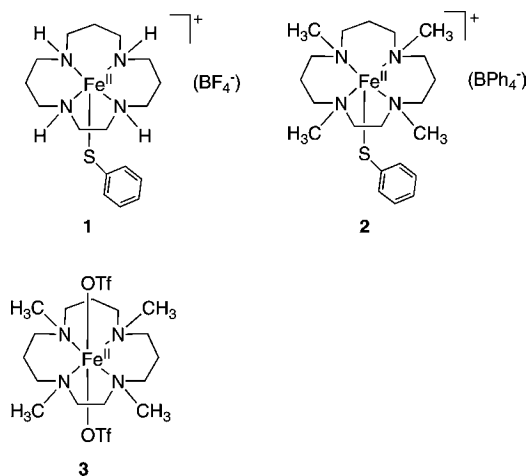
- (10) Pereira, A. S.; Tavares, P.; Folgosa, F.; Almeida, R. M.; Moura, I.; Moura, J. J. G. *Eur. J. Inorg. Chem.* **2007**, 2569–2581.
- (11) Sligar, S. G.; Makris, T. M.; Denisov, I. G. *Biochem. Biophys. Res. Commun.* **2005**, *338*, 346–354.
- (12) Shaik, S.; Kumar, D.; de Visser, S. P.; Altun, A.; Thiel, W. *Chem. Rev.* **2005**, *105*, 2279–2328.
- (13) Loew, G. H.; Harris, D. L. *Chem. Rev.* **2000**, *100*, 407–419.
- (14) Denisov, I. G.; Makris, T. M.; Sligar, S. G.; Schlichting, I. *Chem. Rev.* **2005**, *105*, 2253–2277.

spin Fe^{III} ($S = 1/2$) gives rise to relatively strong Fe–O and weak O–O bonds, while high-spin Fe^{III} ($S = 5/2$) produces weak Fe–O and strong O–O bonds.^{15,16} These studies have led to the hypothesis that a low-spin state in the Fe^{III}–OO(H) intermediate in P450 favors O–O bond cleavage, while an analogous high-spin intermediate in SOR would favor release of H₂O₂.^{17–21} The spin state of the putative Fe^{III}–OO(H) intermediate formed in the native reaction of O₂[–] with SOR is not known, although there is some evidence to suggest that the Fe^{III}–OO(H) species formed upon reaction of SOR with H₂O₂ is high-spin.^{8,22–24} Recent density functional theory (DFT) calculations show that the most energetically favorable reaction pathway for SOR is via a high-spin Fe^{III}–OOH species.³

In addition to spin state, the presence of the cysteine thiolate ligand in SOR may help tune the strength of the Fe–O bond such that Fe–O over O–O bond cleavage occurs preferentially. In a previous study we provided evidence for this idea via the spectroscopic characterization of a series of thiolate-ligated iron model complexes.²⁵ However, it should be noted that a cysteine thiolate is also present in P450, and it has been suggested for many years that the thiolate ligand in P450 promotes O–O bond cleavage via the “push effect.”

The synthesis and characterization of small-molecule model complexes have been critical for gaining an understanding of the spectroscopic and reactivity properties of enzymatic iron-peroxo species. However, there are very few nonheme iron model complexes that mimic the unusual mixed nitrogen/sulfur ligation of SOR, especially those including a thiolate donor in the coordination sphere.^{5,26–29} The lack of such models is in part due to the difficulties inherent in synthesizing complexes with thiolate ligands, which are prone to oxidation as well as forming bridged, polynuclear species. Toward this end we recently synthesized a series of N₄S(thiolate)Fe^{II} complexes with the general formula [Fe^{II}([15]aneN₄)(SAr-*p*-X)]BF₄ ([15]aneN₄ = 1,4,8,12-tetraazacyclopentadecane, X = H (**1**), Cl, OMe, NO₂ and SAr = SC₆F₄-*p*-C₆F₅) as models for the

Chart 1



reduced form of SOR (for the structure of **1**, see Chart 1), in which the electron-donating ability of the thiolate donor was systematically varied.²⁵

These complexes react with alkylhydroperoxides to give metastable, low-spin Fe^{III}–OOR species that can be trapped and spectroscopically characterized at low temperature. Resonance Raman (RR) studies on these complexes showed a clear correlation between an increase in the electron-donating ability of the thiolate ligand and a reduction in the Fe–O stretching frequency. This trend matches nicely with the limited data available for the influence of the Cys donor on the Fe–O stretch in the enzyme.^{6,23}

In the present study we have turned our attention to the influence of the macrocyclic amine nitrogen donors in these model complexes. We hypothesized that alkylation of the nitrogen donors should be a straightforward way of modulating their electronic and steric properties while still allowing us to isolate the same type of coordinatively unsaturated N₄S(thiolate)Fe^{II} model complexes. The alkylation of secondary amine donors in other macrocyclic metal complexes has been shown to influence their redox potentials, LMCT bands, spin state, and reactivity.^{30–32} Although alkylation of the N donors in [15]aneN₄ might be expected to make this ligand a *stronger-field* ligand, previous studies suggested that alkylation should in fact make it a *weaker* donor and perhaps allow us access to *high-spin* Fe^{III}–OO(R or H) species. Only two other *high-spin* thiolate-ligated Fe^{III}–OO(R or H) species have been reported. Kovacs and Solomon³³ described the preparation of high-spin [Fe^{III}(cyclam-PrS)(OOH)]⁺, and Que and Halfen²⁰ reported the high-spin Fe^{III}–OOR complex [Fe^{III}(L⁸py₂)(CH₃-*p*-C₆H₄S)-(OOtBu)]⁺ (L⁸py₂ = neutral N₄ donor).

Herein we describe the synthesis of the new iron(II) complex [Fe^{II}(Me₄[15]aneN₄)(SPh)]BPh₄ (**2**) (Me₄[15]aneN₄ = 1,4,8,12-tetramethyl-1,4,8,12-tetraazacyclopentadecane), which is the tetra-*N*-methylated analogue of [Fe^{II}([15]aneN₄)(SPh)]BF₄ (**1**)

- (15) Lehnert, N.; Ho, R. Y. N.; Que, L., Jr.; Solomon, E. I. *J. Am. Chem. Soc.* **2001**, *123*, 12802–12816.
- (16) Lehnert, N.; Ho, R. Y. N.; Que, L., Jr.; Solomon, E. I. *J. Am. Chem. Soc.* **2001**, *123*, 8271–8290.
- (17) Silaghi-Dumitrescu, R.; Silaghi-Dumitrescu, L.; Coulter, E. D.; Kurtz, D. M., Jr. *Inorg. Chem.* **2003**, *42*, 446–456.
- (18) Roelfes, G.; Vrajmasu, V.; Chen, K.; Ho, R. Y. N.; Rohde, J. U.; Zondervan, C.; la Crois, R. M.; Schudde, E. P.; Lutz, M.; Spek, A. L.; Hage, R.; Feringa, B. L.; Münck, E.; Que, L., Jr. *Inorg. Chem.* **2003**, *42*, 2639–2653.
- (19) Clay, M. D.; Jenney, F. E., Jr.; Hagedoorn, P. L.; George, G. N.; Adams, M. W. W.; Johnson, M. K. *J. Am. Chem. Soc.* **2002**, *124*, 788–805.
- (20) Bukowski, M. R.; Halfen, H. L.; van den Berg, T. A.; Halfen, J. A.; Que, L., Jr. *Angew. Chem., Int. Ed.* **2005**, *44*, 584–587.
- (21) Ogliaio, F.; de Visser, S. P.; Shaik, S. *J. Inorg. Biochem.* **2002**, *91*, 554–567.
- (22) Horner, O.; Mousesca, J. M.; Oddou, J. L.; Jeandey, C.; Nivière, V.; Mattioli, T. A.; Mathé, C.; Fontecave, M.; Maldivi, P.; Bonville, P.; Halfen, J. A.; Latour, J. M. *Biochemistry* **2004**, *43*, 8815–8825.
- (23) Mathé, C.; Nivière, V.; Houee-Levin, C.; Mattioli, T. A. *Biophys. Chem.* **2006**, *119*, 38–48.
- (24) Mathé, C.; Mattioli, T. A.; Horner, O.; Lombard, M.; Latour, J. M.; Fontecave, M.; Nivière, V. *J. Am. Chem. Soc.* **2002**, *124*, 4966–4967.
- (25) Namuswe, F.; Kasper, G. D.; Sarjeant, A. A. N.; Hayashi, T.; Krest, C. M.; Green, M. T.; Moëne-Loccoz, P.; Goldberg, D. P. *J. Am. Chem. Soc.* **2008**, *130*, 14189–14200.
- (26) Brines, L. M.; Kovacs, J. A. *Eur. J. Inorg. Chem.* **2007**, *2*, 9–38.
- (27) Kovacs, J. A. *Chem. Rev.* **2004**, *104*, 825–848.
- (28) Fiedler, A. T.; Halfen, H. L.; Halfen, J. A.; Brunold, T. C. *J. Am. Chem. Soc.* **2005**, *127*, 1675–1689.
- (29) Halfen, J. A.; Moore, H. L.; Fox, D. C. *Inorg. Chem.* **2002**, *41*, 3935–3943.

- (30) Berry, J. F.; Bill, E.; Garcia-Serres, R.; Neese, F.; Weyhermüller, T.; Wieghardt, K. *Inorg. Chem.* **2006**, *45*, 2027–2037.
- (31) Che, C. M.; Wong, K. Y.; Poon, C. K. *Inorg. Chem.* **1986**, *25*, 1809–1813.
- (32) Meyerstein, D. *Coord. Chem. Rev.* **1999**, *186*, 141–147.
- (33) Kitagawa, T.; Dey, A.; Lugo-Mas, P.; Benedict, J. B.; Kaminsky, W.; Solomon, E.; Kovacs, J. A. *J. Am. Chem. Soc.* **2006**, *128*, 14448–14449.

reported previously (Chart 1).^{25,34} Reaction of **2** with *t*BuOOH results in the formation of a novel, metastable Fe^{III}-OOR species that can be trapped at low temperature. Alkylation of the nitrogen donors in the basal plane of this SOR model complex is found to have a profound influence on the spectroscopic (UV-vis, EPR, RR) properties of the alkylperoxo species. In addition, the nonthiolate-ligated complex, Fe^{II}(Me₄[15]aneN₄)(OTf)₂ (**3**) (Chart 1), has been examined and shown to react with ROOH to give a metastable low-temperature Fe^{III}-OOR adduct absent the thiolate donor. Comparison of this species with the thiolate-ligated complex has provided a means to assess the role of the thiolate donor in addition to alkylation at nitrogen.

Experimental Section

General Procedures. All reactions were carried out under an atmosphere of N₂ or Ar using a drybox or standard Schlenk techniques. 1,4,8,12-Tetraazocyclopentadecane ([15]aneN₄) (98%) was purchased from Strem Chemicals. Ferrous triflate was purchased from Wako Chemicals. All other reagents were purchased from commercial vendors and used without further purification unless noted otherwise. Diethyl ether and dichloromethane were purified via a Pure-Solv Solvent Purification System from Innovative Technology, Inc. THF was distilled from sodium/benzophenone. Acetonitrile was distilled from CaH₂. All solvents were degassed by repeated cycles of freeze-pump-thaw and then stored in a drybox. Sodium hydride (60% in mineral oil) was washed with hexanes prior to use. *tert*-Butylhydroperoxide was purchased from Aldrich as an ~5.5 M solution in decane over molecular sieves. Isotopically labeled *t*BuO¹⁸O¹⁸H was synthesized by the reaction of *t*BuMgCl with ¹⁸O₂ following a published procedure.³⁵ The ¹⁸O₂ gas was purchased from Icon Isotopes (99% atom purity).

Physical Methods. Electron paramagnetic resonance (EPR) spectra were obtained on a Bruker EMX EPR spectrometer controlled with a Bruker ER 041 X G microwave bridge at 15 K. The EPR spectrometer was equipped with a continuous-flow liquid He cryostat and an ITC503 temperature controller made by Oxford Instruments, Inc. Low-temperature UV-visible spectra were recorded at -78 °C on a Cary 50 Bio spectrophotometer equipped with a fiber-optic coupler (Varian) and a fiber-optic dip probe (Hellma 661.302-QX-UV, 2 mm path length) for low temperature, using custom-made Schlenk tubes designed for the fiber-optic dip probe. RR spectra were obtained on a custom McPherson 2061/207 spectrometer equipped with a Princeton Instrument liquid-N₂ cooled CCD detector (LN-1100PB). The 647-nm and 514-nm excitations were obtained from a Kr laser (Innova 302, Coherent) and an Ar laser (Innova 90, Coherent), respectively. The laser beam was kept at low power (between 30 and 80 mW) and focused with a cylindrical lens on frozen samples in NMR tubes. The samples were subjected to continuous spinning at 110 K to prevent adverse effects from the laser illumination. Long wave pass filters (RazorEdge filters, Semrock) were used to attenuate the Rayleigh scattering. Sets of ~15 min accumulations were acquired at 4-cm⁻¹ resolution. Frequency calibrations were performed using aspirin and are accurate to ± 1 cm⁻¹. Elemental analyses were performed by Atlantic Microlab Inc., Norcross, GA. ¹H NMR spectra were recorded on a Bruker AMX-400 spectrometer (400 MHz).

Synthesis of 1,4,8,12-Tetramethyl-1,4,8,12-tetraazocyclopentadecane (Me₄[15]aneN₄). The ligand Me₄[15]aneN₄ was synthesized from the reaction of [15]aneN₄ with formaldehyde following a procedure reported by Che et al.³¹ A mixture of [15]aneN₄ (2.0 g, 9.3 mmol), formic acid (9.8 mL, 98–100%), formaldehyde (8.9 mL, 37%), and water (1 mL) was refluxed for 48 h and then cooled

Table 1. Crystallographic Information for Complex **2**

parameter		parameter	
formula	C ₉₃ H ₁₂₄ B ₂ Cl ₆ Fe ₂ N ₈ S ₂	β (deg)	106.1930(10)
<i>M</i> _r	1764.14	γ (deg)	90
cryst syst	monoclinic	<i>V</i> , Å ³	8991.51(19)
space group	<i>P</i> 2 ₁ / <i>c</i>	<i>Z</i>	4
<i>a</i> (Å)	28.3020(4)	cryst color	colorless
<i>b</i> (Å)	18.3435(2)	<i>T</i> , K	110
<i>c</i> (Å)	18.0349(2)	R1 [<i>I</i> > 2σ(<i>I</i>)]	0.0610
α (deg)	90	GOF on <i>F</i> ²	1.046

to room temperature. The reaction mixture was slowly transferred to an 800 mL beaker containing 50 mL of H₂O immersed in an ice bath. A solution of sodium hydroxide (8 M) was gradually added to the mixture until a pH > 12 was reached. The solution was extracted with CH₂Cl₂ (3 × 250 mL). The organic fractions were dried with anhydrous sodium sulfate, and the solvent was evaporated to obtain an oily yellow residue. The oily residue was distilled under reduced pressure (0.1 mmHg) with heating up to 110–120 °C to remove impurities. The pure product remained in the distillation flask as an oil under these conditions. Yield: 2.14 g (85%) of Me₄[15]aneN₄; ¹H NMR (400 MHz, CDCl₃) δ: 1.5, m (CH₂, 6H); 2.2, d (CH₃, 12H); 2.3–2.5, m (CH₂, 16H).

Synthesis of [Fe^{II}(Me₄[15]aneN₄)(SC₆H₅)₂](BPh₄) (2**).** A solution of Me₄[15]aneN₄ (0.135 g, 0.498 mmol) in CH₃CN (3 mL) was added dropwise to a suspension of anhydrous FeCl₂ (0.053 g, 0.421 mmol) in CH₃CN (3 mL) and stirred for 45 min resulting in a pale yellow-brown solution. An acetonitrile solution of NaSC₆H₅ (0.089 g, 0.676 mmol) (prepared from addition of benzenethiol to NaH in THF) was added to the mixture dropwise and stirred for 1 h to give a brown solution with a white precipitate. An acetonitrile solution of NaBPh₄ (0.157 g, 0.460 mmol) was added to the mixture dropwise, resulting in more white precipitate, and the solution was stirred overnight. The solvent was removed under vacuum to give an off-white solid, which was redissolved in CH₂Cl₂ and filtered through Celite to give a brown solution. Vapor diffusion of diethyl ether into this solution resulted in colorless plates of **2** after a few hours. Yield: 0.260 g (78%); Anal. Calcd for C_{45.5}H₆₀N₄BClFeS (2·0.5 CH₂Cl₂) C, 68.55; H, 7.59; N, 7.03. Found: C, 68.54; H, 7.74; N, 6.97.

Synthesis of Fe^{II}(Me₄[15]aneN₄)(OTf)₂ (3**).** The synthesis of **3** was reported previously,³⁶ and this method was followed with a slight modification. To a suspension of anhydrous Fe(OTf)₂ (0.169 g, 0.477 mmol) in CH₃CN (3 mL) was added a solution of Me₄[15]aneN₄ (0.155 g, 0.572 mmol) in CH₃CN (3 mL) dropwise, and the mixture was stirred for 5 h resulting in a gray-brown suspension. Acetonitrile was removed under vacuum to give a brown solid which was redissolved in CH₂Cl₂ and filtered through Celite to give a clear solution. Vapor diffusion of diethyl ether into this solution resulted in crystals (colorless plates) of **3** within 24 h. Yield: 0.222 g (75%); Anal. Calcd for C₁₇H₃₄F₆FeN₄O₆S₂: C, 32.70; H, 5.49; N, 8.97. Found: C, 32.78; H, 5.49; N, 8.93. A unit cell check on representative crystals of **3** were performed by X-ray diffraction and compared with that reported previously,³⁶ confirming the structure of **3**.

X-ray Crystallography of 2. X-ray diffraction data were collected on an Oxford Diffraction Xcalibur3 diffractometer, equipped with a Mo Kα sealed-tube source and a Sapphire CCD detector. Single crystals were mounted in oil under a cold nitrogen stream at 110(2) K. Selected crystallographic details are provided in Table 1. Data were collected, integrated, scaled, and corrected for absorption using CrysAlis PRO software.³⁷ Structures were solved and refined using the SHELXTL suite of software.³⁸

Each unit cell contains two independent iron molecules and three methylene chloride solvent molecules, i.e. 1.5 CH₂Cl₂ per Fe^{II}. The

(34) Krishnamurthy, D.; Kasper, G. D.; Namuswe, F.; Kerber, W. D.; Sarjeant, A. A. N.; Moëgne-Loccoz, P.; Goldberg, D. P. *J. Am. Chem. Soc.* **2006**, *128*, 14222–14223.

(35) Walling, C.; Buckler, S. A. *J. Am. Chem. Soc.* **1955**, *77*, 6032–6038.

(36) Suh, Y.; Seo, M. S.; Kim, K. M.; Kim, Y. S.; Jang, H. G.; Tosha, T.; Kitagawa, T.; Kim, J.; Nam, W. *J. Inorg. Biochem.* **2006**, *100*, 627–633.

cationic portions of the two molecules are disordered across the pseudomirror plane of the $\text{Me}_4[15]\text{aneN}_4$ ligand such that a CH_2CH_2 group of the ligand in each cation exchanges positions with a $\text{CH}_2\text{CH}_2\text{CH}_2$ group. There is also some disorder in one methylene chloride molecule.

Electrochemistry. Cyclic voltammetry was performed on an EG&G Princeton Applied Research potentiostat/galvanostat model 263A at scan rates of 0.025–0.5 V/s. A three-electrode configuration made up of a platinum working electrode, a Ag/AgCl reference electrode (3.5 M KCl), and a platinum wire auxiliary electrode was employed. Measurements were performed with 1.0 mM analyte in CH_2Cl_2 at ambient temperatures under argon using 0.10 M tetra-*n*-butylammonium hexafluorophosphate (recrystallized 3 times from ethanol/water, dried under high vacuum for 3 days, and stored in the drybox) as the supporting electrolyte. The ferrocenium/ferrocene couple ($\text{FeCp}_2^{+/0}$) at 0.45 V was used as an external reference.

Generation of $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{SPh})(\text{OO}t\text{Bu})]^+$ (2a**) for UV–vis Spectroscopy.** In a typical reaction, a custom-made Schlenk flask was loaded with 8–12 mg of $[\text{Fe}^{\text{II}}(\text{Me}_4[15]\text{aneN}_4)(\text{SPh})]\text{BPh}_4$ (**2**) which was dissolved in 5 mL of CH_2Cl_2 to give a concentration of 1–3 mM for the Fe^{II} complex. A fiber-optic dip probe UV–vis cell with a path length of 2 mm was inserted into the flask, and the solution was cooled to -78°C . After the UV–vis spectrum of the Fe^{II} starting material was recorded, 1.75 equiv of *t*BuOOH (stock solution in *n*-decane) diluted in CH_2Cl_2 was added to the Fe^{II} complex. UV–vis spectra were recorded immediately following the addition of *t*BuOOH. An instant color change from colorless to turquoise green was observed, and the formation and decay of the turquoise species were monitored by UV–vis spectroscopy.

Generation of $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{OTf})(\text{OO}t\text{Bu})]^+$ (3a**) for UV–vis Spectroscopy.** In a typical reaction, a custom-made Schlenk flask was loaded with $[\text{Fe}^{\text{II}}(\text{Me}_4[15]\text{aneN}_4)(\text{OTf})_2$ (**3**) (11 mg, 0.0176 mmol) which was then dissolved in CH_2Cl_2 (5 mL, 3.5 mM in **3**). A fiber-optic dip probe UV–vis cell with a path length of 2 mm was inserted into the flask, and the solution was cooled to -23°C . After recording the UV–vis spectrum of the Fe^{II} starting material, 1.75 equiv of *t*BuOOH (stock solution in *n*-decane) diluted in CH_2Cl_2 was added to the Fe^{II} complex. UV–vis spectra were recorded immediately following the addition of *t*BuOOH. A gradual color change from colorless to dark blue was observed, and the formation and decay of the new species were monitored by UV–vis spectroscopy.

Generation of $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{SPh})(\text{OO}t\text{Bu})]^+$ (2a**) for Resonance Raman Spectroscopy.** In a typical procedure, a Wilmad WG-5 M economy NMR tube was loaded with a solution of **2** in CH_2Cl_2 (300 μL , 19 mM) and cooled to -78°C . An amount of *t*BuOOH (1.75 equiv, stock solution in *n*-decane) diluted in CH_2Cl_2 was added to the NMR tube. The reaction mixture was shaken carefully to obtain a homogeneous solution, and the rapid evolution of a turquoise color was noted, indicating the formation of **2a**. The solution was then frozen in liquid nitrogen and stored at 77 K until RR measurements were taken. Samples containing $t\text{Bu}^{18}\text{O}^{18}\text{OH}$ were prepared in an identical manner.

Generation of $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{OTf})(\text{OO}t\text{Bu})]^+$ (3a**) for Resonance Raman Spectroscopy.** In a typical procedure, a Wilmad WG-5 M economy NMR tube was loaded with a solution of **3** in CH_2Cl_2 (300 μL , 20 mM) and cooled to -23°C . An amount of *t*BuOOH (1.75 equiv, stock solution in *n*-decane) diluted in CH_2Cl_2 was added to the NMR tube. The reaction mixture was shaken to obtain a homogeneous solution, and the evolution of a dark blue color was observed, indicating the formation of **3a**. The solution was then frozen in liquid nitrogen and stored at 77 K until

RR measurements were taken. Samples containing $t\text{Bu}^{18}\text{O}^{18}\text{OH}$ were prepared in an identical manner.

Generation of $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{SPh})(\text{OO}t\text{Bu})]^+$ (2a**) for EPR Spectroscopy.** In a typical procedure, a Wilmad Quartz EPR tube was loaded with a solution of **2** in CH_2Cl_2 (400 μL , 3 mM) and cooled to -78°C . An amount of *t*BuOOH (1.75 equiv, stock solution in *n*-decane) diluted in CH_2Cl_2 was added to the EPR tube. The reaction mixture was bubbled briefly with Ar gas to obtain a homogeneous solution, taking care so as to minimize warming of the solution. The rapid evolution of a turquoise color was noted, indicating the formation of **2a**. The solution was then frozen in liquid nitrogen and stored at 77 K until EPR measurements were taken.

Generation of $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{OTf})(\text{OO}t\text{Bu})]^+$ (3a**) for EPR Spectroscopy.** In a typical procedure, a Wilmad Quartz EPR tube was loaded with a solution of **3** in CH_2Cl_2 (400 μL , 5 mM) and cooled to -23°C . An amount of *t*BuOOH (1.75 equiv, stock solution in *n*-decane) diluted in CH_2Cl_2 was added to the EPR tube. The reaction mixture was bubbled briefly with Ar gas to obtain a homogeneous solution, taking care so as to minimize warming of the solution. The evolution of a dark blue color was noted, indicating the formation of **3a**. The solution was then frozen in liquid nitrogen and stored at 77 K until EPR measurements were taken.

Results and Discussion

Selection of the Tetramethylated Ligand $\text{Me}_4[15]\text{aneN}_4$. Our previous work on models of the active site of SOR employed the 15-membered macrocycle $[15]\text{aneN}_4$. This macrocyclic, tetradentate nitrogen donor was selected because it provided the appropriate number of neutral N donors to mimic the four His donors in SOR, was commercially available, and, like other macrocyclic ligands, was known to give very stable complexes. Although there were no structurally characterized iron(III) complexes of $[15]\text{aneN}_4$, a variety of other trivalent complexes of other metals (Co^{III} , Rh^{III} , Cr^{III}) had been described previously.^{39–42} These trivalent complexes all contain *trans*-ligated axial ligands (*trans*- $([15]\text{aneN}_4)\text{M}^{\text{III}}(\text{L})_2$), despite the potential flexibility of macrocycles of this type to fold into a *cis*-ligated structure, and this propensity for forming *trans* structures was desirable from the standpoint of preparing *trans*-ligated iron(III) thiolate/peroxo species. In this study we employed the tetramethylated analogue of $\text{Me}_4[15]\text{aneN}_4$, which was expected to offer the same advantages as those outlined above for $[15]\text{aneN}_4$.

Synthesis and Characterization of Iron(II) Complexes. The iron(II) complex $[\text{Fe}^{\text{II}}(\text{Me}_4[15]\text{aneN}_4)(\text{SPh})]\text{BPh}_4$ (**2**) was prepared according to Scheme 1. Successful synthesis of **2** requires the use of an anhydrous metal salt in combination with dry, aprotic solvents. Attempts to synthesize **2** from hydrated metal salts such as $\text{Fe}(\text{BF}_4)_2 \cdot 6\text{H}_2\text{O}$, or protic solvents such as MeOH, were unsuccessful. It was previously shown by Halfen and co-workers that the synthesis of the related cyclam complex $[\text{Fe}^{\text{II}}(\text{Me}_4\text{cyclam})(\text{SC}_6\text{H}_4\text{-}p\text{-OMe})]\text{OTf}$ requires the use of rigorously dry, aprotic conditions, such as employing the specially prepared triflate starting material, $\text{Fe}(\text{OTf})_2 \cdot \text{CH}_3\text{CN}$, in dry THF. It was suggested that these conditions were necessary to avoid protonation of the tetraamine macrocycle.²⁸

As shown in Scheme 1, the reaction of anhydrous FeCl_2 with $\text{Me}_4[15]\text{aneN}_4$ in dry CH_3CN results in the insertion of Fe^{II} into

(37) *CrysAlis PRO*; Oxford Diffraction Ltd.: Wroclaw, Poland, 2008.

(38) SHELDRICK, G. M. *SHELXTL*, version 610; Bruker AXS Inc.: Madison, Wisconsin, USA.

(39) Bhattacharya, P. K. *Dalton Trans.* **1980**, 810–812.

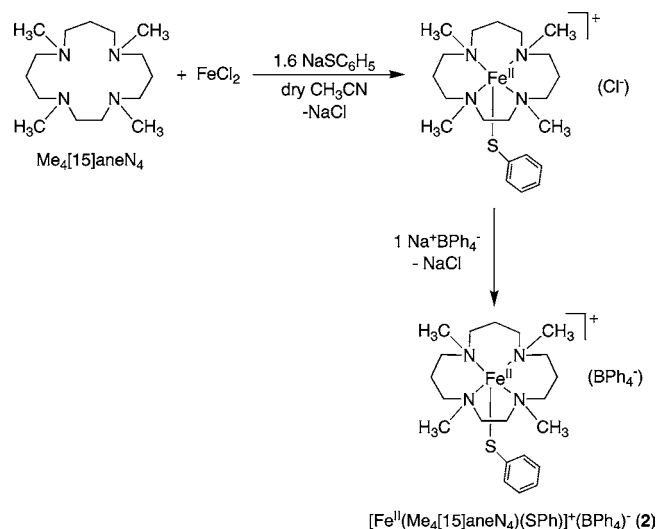
(40) Hay, R. W.; Tarafder, M. T. H. *Dalton Trans.* **1991**, 823–827.

(41) Islam, M. S.; Uddin, M. M. *Polyhedron* **1993**, *12*, 423–426.

(42) Hung, Y.; Busch, D. H. *J. Am. Chem. Soc.* **1977**, *99*, 4977–4984.

(43) Addison, A. W.; Rao, T. N.; Reedijk, J.; van Rijn, J.; Verschoor, G. C. *Dalton Trans.* **1984**, 1349–1456.

Scheme 1



the Me₄[15]aneN₄ cavity, as evidenced by the fact that FeCl₂, which is sparingly soluble in CH₃CN, gradually dissolves upon addition of Me₄[15]aneN₄ to give a yellow-brown solution. The thiolate donor is then added as a solution of the sodium salt (NaSC₆H₅) in CH₃CN, leading to the formation of a dark brown solution and a white precipitate, presumably NaCl. As found for the syntheses of the thiolate-ligated complexes [(Fe^{II}([15]aneN₄)(SAr))⁺,²⁵ the preparation of [Fe^{II}(Me₄[15]aneN₄)(SPh)]⁺ was quite sensitive to the amount of arylthiolate ligand used. It was found that addition of a slight excess of the thiolate ligand (1.6 equiv) was optimal for obtaining **2** in good yield as a crystalline solid. Crystals of **2** were obtained via metathesis of the chloride anion with the noncoordinating, bulky tetraphenylborate anion, followed by recrystallization from CH₂Cl₂/Et₂O. This preparation was used to prepare X-ray quality crystals as well as for routine preparation of the complex for all other studies.

We prepared the known, nonthiolate ligated iron(II) complex [Fe^{II}(Me₄[15]aneN₄)(OTf)₂] (**3**) to examine a close analogue of **2** that did not contain a thiolate donor, thereby giving us a direct means of determining the influence of the thiolate donor on the structure and reactivity of the iron(II) center. The synthesis of **3** from Fe(OTf)₂ and Me₄[15]aneN₄ was reported previously,³⁶ and we followed this same method except for the recrystallization procedure, which involved vapor diffusion of Et₂O directly into reaction mixtures of **3**. We found that higher yields of **3** could be obtained by vapor diffusion of Et₂O into solutions of **3** in CH₂Cl₂.

X-ray Structural Studies. X-ray quality crystals of **2** were grown by vapor diffusion of Et₂O into a methylene chloride solution of **2**. The crystal structure of the cation of **2** is shown as an ORTEP diagram in Figure 1. Structural parameters are shown in Table 1, and selected bond lengths and angles are given in Table 2. There are two independent molecules of **2** in the unit cell, both of which are shown in Figure 1. The iron(II) center in each molecule is five-coordinate, with four nitrogen donors from the Me₄[15]aneN₄ ligand and a sulfur donor from the PhS⁻ ligand completing the coordination sphere.

Both molecules exhibit distorted square pyramidal geometries ($\tau^{43} = 0.31$; square pyramid = $0 < \tau < 1.0$ = trigonal bipyramid) and differ mainly in the relative position of the phenylthiolate ligand. In molecule 1 (Fe1), the phenyl ring of the PhS⁻ group sits directly over N2, such that the torsion angle defined by N2–Fe1–S1–C(Ph) is nearly zero ($-4.55(17)^\circ$). In molecule

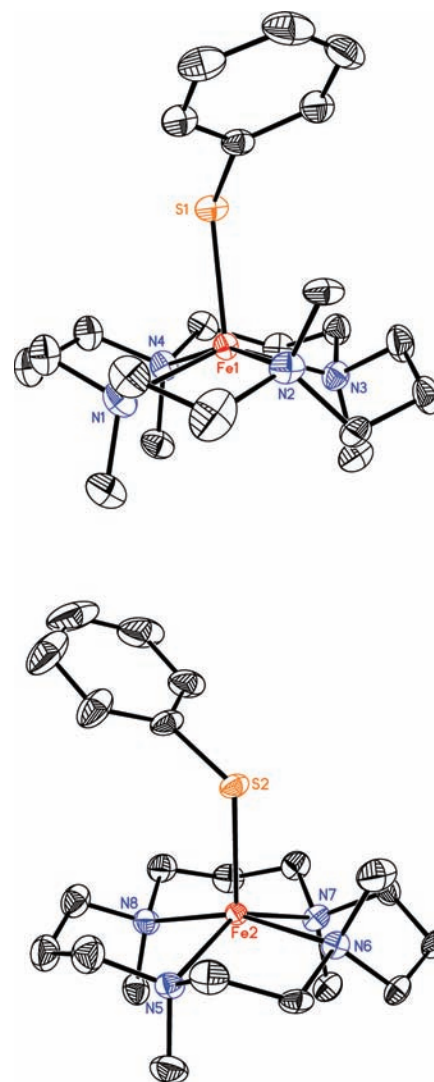


Figure 1. ORTEP diagrams of the two independent complex cations of **2** (molecule 1, top; molecule 2, bottom) showing 30% probability thermal ellipsoids. Hydrogen atoms were omitted for clarity.

Table 2. Selected Bond Distances (Å) and Angles (deg) for Complex **2**

Fe1–N1	2.185(2)	N1–Fe1–N2	79.90(13)
Fe1–N2	2.340(5) ^a	N1–Fe1–N3	143.48(10)
	2.386(16) ^b		
Fe1–N3	2.217(2)	N1–Fe1–N4	88.48(9)
Fe1–N4	2.267(2)	N3–Fe1–N4	90.51(9)
Fe1–S1	2.3540(8)	N2–Fe1–N3	90.96(13)
Fe2–N5	2.157(2)	N2–Fe1–N4	162.28(13)
Fe2–N6	2.406(5) ^a	N5–Fe2–N6	77.94(13)
	2.370(8) ^b		
Fe2–N7	2.177(2)	N5–Fe2–N7	143.47(9)
Fe2–N8	2.247(2)	N5–Fe–N8	90.29(9)
Fe2–S2	2.3459(7)	N6–Fe2–N7	91.34(13)
		N6–Fe2–N8	163.30(14)
		N7–Fe2–N8	91.21(9)

^a Major component in which the CH₂CH₂ group of the ligand is in one position. ^b Minor component in which CH₂CH₂ group of the ligand is in another position.

2, the phenyl ring of the thiolate ligand is rotated almost 180° in relation to molecule 1, being positioned directly over N8 and resulting in a torsion angle of N6–Fe2–S2–C(Ph) = $-171.99(18)^\circ$. An ethyl bridge between N1 and N2 (molec 1) and N5 and N6

(molec 2) is positionally disordered with an adjacent propyl bridge, which was well modeled in the final refinements.

The four methylated nitrogen atoms are chiral, and as a consequence there are six possible diastereomers that can be characterized by the positions of the methyl groups in relation to the plane of the N donors. The diastereomer that is seen in Figure 1 is the “3-down-1-up” (*trans*-I⁴⁰) configuration, in which one N–CH₃ group (N(2)–CH₃ and N(6)–CH₃) is on the same side of the macrocyclic plane as the thiolate ligand, while the other three N–CH₃ groups lie on the opposite side. This is the same as that observed previously for the N–H groups in [Fe^{II}([15]aneN₄)(SAR)]BF₄.²⁵ In comparison, the six-coordinate complex [Fe^{II}(Me₄[15]aneN₄)(OTf)₂ (**3**) is found in a 2-up-2-down configuration (*trans*-II).³⁶ This configuration likely helps reduce the steric interactions between the two axial triflate ligands and the N–CH₃ groups. Similarly, in five-coordinate **2** the 3-down-1-up configuration helps to minimize steric repulsion between the phenylthiolate ligand and the N-Me substituents. To our knowledge there are no other structurally characterized Me₄[15]aneN₄ complexes with the exception of [Ru(Me₄[15]aneN₄)(O)₂](ClO₄)₂.⁴⁴ There are some structures of the 14-membered analogue, such as [Ru(Me₄[14]aneN₄)(O)(Cl)]⁺, which is six-coordinate like complex **3** but found in the “3-up-1-down” configuration, or [Ru(Me₄[14]aneN₄)(O)₂]⁺, which is found in the “4-up” configuration. Both [Ru^{IV}(Me₄[14]aneN₄)(O)(NCCH₃)]²⁺⁴⁵ and [Fe^{IV}(Me₄[14]aneN₄)(O)(NCCH₃)]²⁺⁴⁶ have been crystallographically characterized and are also found in the “4-up” configuration, where the four methyl groups point toward the NCCH₃ ligand. The six-coordinate complexes therefore do not necessarily favor a 2-up-2-down configuration as seen for the bis-triflate complex **3**.

The average Fe–N distances for **2** are 2.2410 and 2.2468 Å for molecules 1 and 2, respectively, and fall in line with other high-spin Fe^{II} complexes, including complex **3**³⁶ (Fe–N_{AVE} = 2.2445 Å). These bonds are slightly elongated compared to the average Fe–N distance of 2.1953 Å for the nonmethylated complex [Fe^{II}([15]aneN₄)(SPh)]BF₄. The Fe–S bond distances of 2.3540(8) and 2.3459(7) Å for molecules 1 and 2, respectively, are typical for high-spin Fe^{II}–SR complexes^{47–49} and are similar to the Fe–S bond distance for [Fe^{II}([15]aneN₄)(SPh)]BF₄. The Fe–N and Fe–S distances are also similar to those found in the reduced (Fe^I) form of SOR for the His and Cys donors,^{8,50–52} and thus complex **2** is a good structural model of this site.

Methylation of the secondary amines in [15]aneN₄ make it a weaker-field ligand. This weakening is evident in the slight elongation of the average Fe–N distance for **2** as compared to [Fe^{II}([15]aneN₄)(SPh)]BF₄. This result is in line with previous work from Meyerstein^{53,54} and Wiegardt,³⁰ which showed that alkylation of the amine donors in this kind of macrocyclic ligand

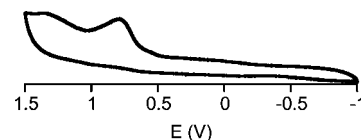


Figure 2. Cyclic voltammogram for [Fe^{II}(Me₄[15]aneN₄)(SPh)]BPh₄ in CH₂Cl₂ using a Ag/AgCl reference electrode (3.5 M KCl).

should result in a *weaker-field* ligand. DFT calculations also support this conclusion.³⁰

Electrochemistry. The cyclic voltammogram for **2** is shown in Figure 2. An irreversible oxidation wave is observed, with an anodic potential of +780 mV (vs Ag/AgCl), which we assign to the Fe^{III/II} couple. In comparison, [Fe^{II}([15]aneN₄)(SPh)]BF₄ (**1**) exhibits an Fe^{III/II} potential at +501 mV.²⁵ Thus alkylation of the amine donors makes the complex more difficult to oxidize by 279 mV, supporting the conclusion that alkylation of the nitrogen atoms in Me₄[15]aneN₄ makes it a much weaker donor than the nonalkylated [15]aneN₄. Solvation factors have been invoked to account for the effect of N-methylation on the redox potentials of nickel(II/I) and chromium(III/II) cyclam complexes^{32,55} and may also contribute to the difference in redox potentials between **1** and **2**.

The redox potential for the enzyme SOR has been measured by different methods, giving a range of 200–365 mV for the Fe^{III/II} couple.^{19,56–58} The oxidation of **2** is clearly at a much higher potential than that of the enzyme, as has been found for other SOR models from our group and others.²⁵ Multiple factors are likely to contribute to the differences in redox potentials between the enzyme and model complexes (e.g., aqueous vs nonaqueous conditions, H-bonding).

Reaction of Fe^{II} Complexes with Alkylhydroperoxides (ROOH). Addition of *t*BuOOH to **2** in CH₂Cl₂ at –78 °C leads to a rapid color change from colorless for the iron(II) complex to a turquoise green characteristic of a metastable species **2a** (Scheme 2). The new turquoise species gives rise to a UV–vis spectrum with λ_{max} at 340, 405, and 650 nm (Figure 3). The peak at 650 nm (ε = 2300 M^{–1} cm^{–1}) is assigned as an alkylperoxo-to-iron(III) LMCT band based on both the energy of this transition and its molar absorptivity.^{16,59–61} There is an

(44) Mak, T. C. W.; Che, C. M.; Wong, K. Y. *Chem. Commun.* **1985**, 986–988.

(45) Che, C. M.; Wong, K. Y.; Mak, T. C. W. *Chem. Commun.* **1985**, 546–548.

(46) Rohde, J. U.; In, J. H.; Lim, M. H.; Brennessel, W. W.; Bukowski, M. R.; Stubna, A.; Münck, E.; Nam, W.; Que, L., Jr. *Science* **2003**, 299, 1037–1039.

(47) Krishnamurthy, D.; Sarjeant, A. N.; Goldberg, D. P.; Caneschi, A.; Totti, F.; Zakharov, L. N.; Rheingold, A. L. *Chem.–Eur. J.* **2005**, 11, 7328–7341.

(48) Mukherjee, R. N.; Abrahamson, A. J.; Patterson, G. S.; Stack, T. D. P.; Holm, R. H. *Inorg. Chem.* **1988**, 27, 2137–2144.

(49) Zang, Y.; Que, L., Jr. *Inorg. Chem.* **1995**, 34, 1030–1035.

(50) Adam, V.; Royant, A.; Nivière, V.; Molina-Heredia, F. P.; Bourgeois, D. *Structure* **2004**, 12, 1729–1740.

(51) Santos-Silva, T.; Trincao, J.; Carvalho, A. L.; Bonifacio, C.; Auchere, F.; Raleiras, P.; Moura, I.; Moura, J. J. G.; Romao, M. J. *J. Biol. Inorg. Chem.* **2006**, 11, 548–558.

(52) Yeh, A. P.; Hu, Y. L.; Jenney, F. E., Jr.; Adams, M. W. W.; Rees, D. C. *Biochemistry* **2000**, 39, 2499–2508.

(53) Golub, G.; Cohen, H.; Paoletti, P.; Bencini, A.; Meyerstein, D. *Dalton Trans.* **1996**, 2055–2060.

(54) Golub, G.; Zilbermann, I.; Cohen, H.; Meyerstein, D. *Supramol. Chem.* **1996**, 6, 275–279.

(55) Clark, T.; Hennemann, M.; van Eldik, R.; Meyerstein, D. *Inorg. Chem.* **2002**, 41, 2927–2935.

(56) Jovanovic, T.; Ascenso, C.; Hazlett, K. R. O.; Sikkink, R.; Krebs, C.; Litwiller, R.; Benson, L. M.; Moura, I.; Moura, J. J. G.; Radolf, J. D.; Huynh, B. H.; Naylor, S.; Rusnak, F. *J. Biol. Chem.* **2000**, 275, 28439–28448.

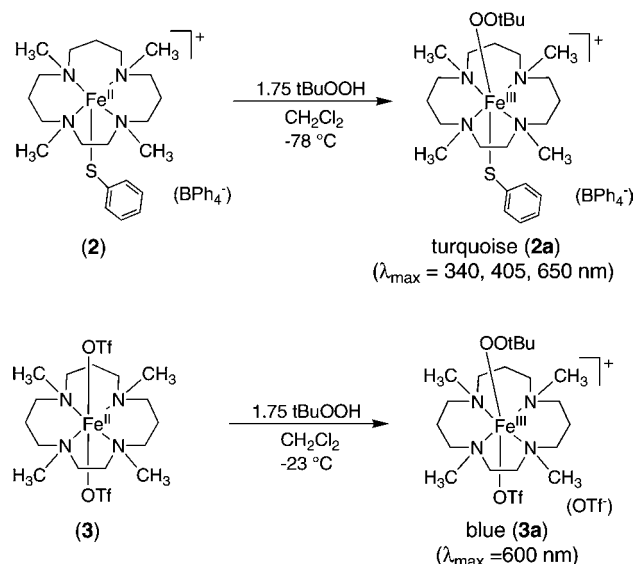
(57) Rodrigues, J. V.; Saraiva, L. M.; Abreu, I. A.; Teixeira, M.; Cabelli, D. E. *J. Biol. Inorg. Chem.* **2007**, 12, 248–256.

(58) Tavares, P.; Ravi, N.; Moura, J. J. G.; LeGall, J.; Huang, Y. H.; Crouse, B. R.; Johnson, M. K.; Huynh, B. H.; Moura, I. *J. Biol. Chem.* **1994**, 269, 10504–10510.

(59) Jensen, M. P.; Costas, M.; Ho, R. Y. N.; Kaizer, J.; Payeras, A. M. I.; Münck, E.; Que, L., Jr.; Rohde, J. U.; Stubna, A. *J. Am. Chem. Soc.* **2005**, 127, 10512–10525.

(60) Jensen, M. P.; Payeras, A. M. I.; Fiedler, A. T.; Costas, M.; Kaizer, J.; Stubna, A.; Münck, E.; Que, L., Jr. *Inorg. Chem.* **2007**, 46, 2398–2408.

Scheme 2



intense feature in the spectrum of the Fe^{II} complex **2** at $\lambda_{\max} = 313 \text{ nm}$ that can be assigned to a S \rightarrow Fe^{II} charge-transfer band based on a detailed analysis of a series of closely related iron(II)–thiolate complexes.²⁸ This band apparently shifts to 340 nm upon formation of **2a**. A number of thiolate-to-iron(III) CT bands have been assigned previously by spectroscopic and computational methods, although bands in the high-energy region ($\lambda_{\max} < 400 \text{ nm}$) are more difficult to identify exclusively as S \rightarrow Fe^{III} CT bands because of a possible overlap with other transitions.^{19,62} We suggest that the peak at 340 nm seen for **2a** is a S \rightarrow Fe^{III} CT band, and the fact that this peak is absent in the triflate-derived alkylperoxo-iron(III) complex (vide infra) supports this assignment. However, this assignment must remain tentative until a more detailed spectroscopic and/or theoretical analysis can be carried out. The shoulder at 405 nm grows in together with the 340 and 650 nm bands, clearly arising from **2a** as well. The mechanism of formation of the alkylperoxo species likely follows that proposed previously for the non-methylated complexes, occurring in two steps. The initial step involves conversion of the Fe^{II} complex to an Fe^{III} complex, presumably Fe^{III}–OH, via oxidation with 0.5 equiv of ROOH. This step is then followed by displacement of hydroxide by ROOH to give Fe^{III}–OOR. Our earlier work on the nonmethylated analogues provided evidence for such a mechanism from titration studies, and Que and co-workers have provided data in support of this mechanism for the formation of other Fe–OOR(H) complexes from Fe^{II} + excess ROOH or H₂O₂.^{18,63}

The spectrum for the turquoise species evolves over $\sim 40 \text{ min}$ and then gradually decays with a similar rate to a purple species with a broad absorption centered at 514 nm (Figure 3). This purple species is stable for several hours at low temperature but decays rapidly upon warming to room temperature. The broad absorption at 514 nm is consistent with an alkylperoxo-to-iron(III) LMCT band, but this spectrum lacks the 340 nm feature observed in **2a**. These data suggest that the thiolate ligand may be lost in the purple species (vide infra). For the

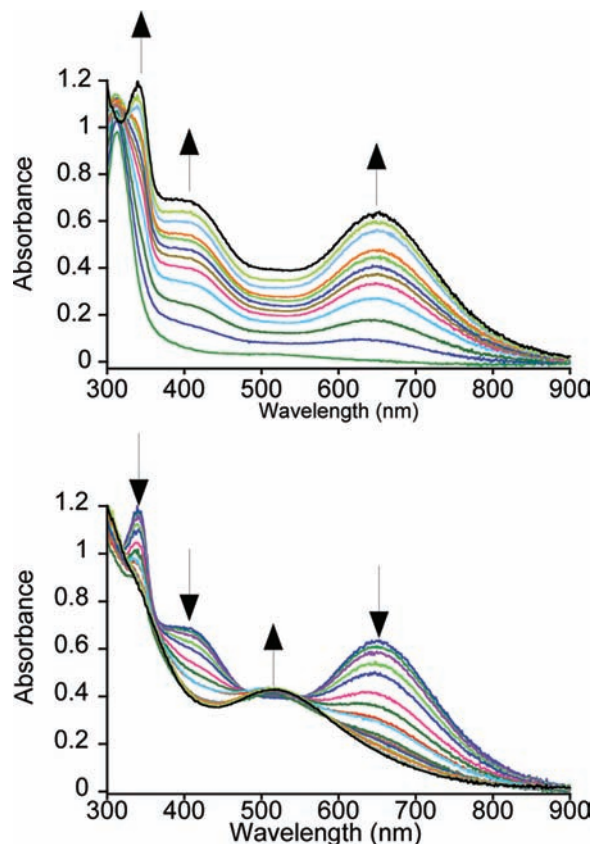


Figure 3. UV–vis spectral changes for the reaction of [Fe^{II}(Me₄[15]aneN₄)(SPh)](BPh₄) and *t*BuOOH in CH₂Cl₂ at $-78 \text{ }^\circ\text{C}$, showing the formation (top) and decay (bottom) of [Fe^{III}(Me₄[15]aneN₄)(SPh)(OO*t*Bu)]⁺ (**2a**). The formation was monitored every 30 s for 45 min, and the decay was monitored every 2–5 min for 40 min (select spectra shown for clarity).

nonmethylated analogues [Fe^{III}([15]aneN₄)(SAr)(OOR)]⁺, it was shown that the thiolate ligand is converted to disulfide upon decomposition of the alkylperoxo-iron(III) complex at low temperature.²⁵

The UV–vis spectrum for the nonmethylated [Fe^{III}([15]aneN₄)(SPh)(OO*t*Bu)]⁺ exhibits an alkylperoxo-to-iron(III) LMCT band at 526 nm, corresponding to a deep burgundy-colored intermediate, whereas the tetramethylated complex **2a** exhibits a strongly red-shifted absorption at 650 nm associated with the turquoise chromophore. Thus there is a strong influence of N-methylation on the alkylperoxo-to-iron(III) LMCT band, and previous work from others supports this notion.^{53,64–68} For example, Meyerstein and co-workers observed a red shift in the N-to-Cu^{II} LMCT band of a Cu^{II} complex upon methylation of a linear polyamine ligand.⁵³ The lowering in energy of the alkylperoxo-to-iron(III) band for **2a** also tracks with the increase in the oxidation potential for this complex versus the unmethylated analogue. It is reasonable to assume that the energy of the alkylperoxide orbital involved in the LMCT transition remains

(61) Ménage, S.; Wilkinson, E. C.; Que, L., Jr.; Fontecave, M. *Angew. Chem., Int. Ed.* **1995**, *34*, 203–205.

(62) Kennepohl, P.; Neese, F.; Schweitzer, D.; Jackson, H. L.; Kovacs, J. A.; Solomon, E. I. *Inorg. Chem.* **2005**, *44*, 1826–1836.

(63) Kim, J.; Zang, Y.; Costas, M.; Harrison, R. G.; Wilkinson, E. C.; Que, L., Jr. *J. Biol. Inorg. Chem.* **2001**, *6*, 275–284.

(64) Bazzicalupi, C.; Bencini, A.; Cohen, H.; Giorgi, C.; Golub, G.; Meyerstein, D.; Navon, N.; Paoletti, P.; Valtancoli, B. *Dalton Trans.* **1998**, 1625–1631.

(65) Bernhardt, P. V. *J. Am. Chem. Soc.* **1997**, *119*, 771–774.

(66) Golub, G.; Cohen, H.; Paoletti, P.; Bencini, A.; Messori, L.; Bertini, I.; Meyerstein, D. *J. Am. Chem. Soc.* **1995**, *117*, 8353–8361.

(67) Jubran, N.; Ginzburg, G.; Cohen, H.; Koresch, Y.; Meyerstein, D. *Inorg. Chem.* **1985**, *24*, 251–258.

(68) Wagner, F.; Barefield, E. K. *Inorg. Chem.* **1976**, *15*, 408–417.

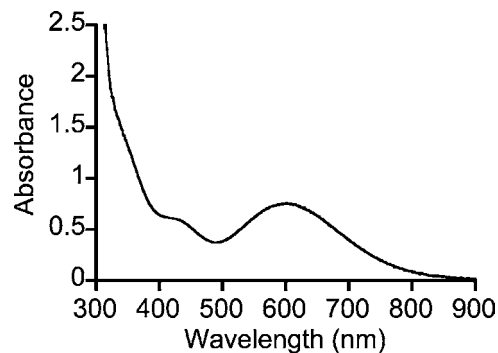


Figure 4. UV-vis spectrum of $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{OTf})(\text{OOtBu})]^+$ (**3a**) in CH_2Cl_2 at -23°C .

essentially the same for **2a** and its unmethylated analogue. The increase in the oxidation potential for **2** suggests a stabilization of the HOMO, probably dominated by metal *d* orbital character, which also results in a lowering of the energy of the LMCT transition in the alkylperoxo complex. This analysis, however, does not take into account the spin-state change that occurs on going from the unmethylated analogue to **2a** (vide infra). The same correlation for a series of $\text{Fe}^{\text{III}}\text{--OOH}$ complexes has been made, where an increase in the $\text{Fe}^{\text{III/II}}$ potential of iron(II) precursors was found to be linearly dependent on a decrease in the energy of the hydroperoxo-to-iron(III) LMCT absorptions.¹⁸

The nonthiolate-ligated complex **3** does not react with *t*BuOOH at -78°C , but once the temperature is raised to -40°C , a faint blue color develops. Moreover, when the reaction is run at -23°C , a gradual color change takes place from colorless Fe^{II} to dark blue (Scheme 2). Low-temperature UV-vis reveals a peak with λ_{max} 600 nm ($\epsilon = 1080 \text{ M}^{-1} \text{ cm}^{-1}$), indicative of an alkylperoxo-to-iron(III) LMCT band (Figure 4). This new blue species, **3a**, forms over 60 min and then decays with a similar rate to give a yellow solution. The appearance of a second chromophore in the decay of **3a** was not observed (Figure S1), in contrast to what was seen for the decay of **2a**. The necessity to employ higher temperatures in the reaction of **3** may be a consequence of the lack of an open site on the metal, hindering the initial coordination and oxidation by ROOH.

EPR Spectroscopy. The X-band EPR spectra for **2a** and **3a** in CH_2Cl_2 at 15 K are shown in Figure 5, and the *g* values are collected in Table 3 with other literature values for comparison. The spectra are typical of high-spin ($S = 5/2$) iron(III) complexes. The turquoise complex **2a** (Figure 5, top) exhibits a sharp, intense peak at *g* 4.3 that extends out on the high-field side by over 400 G. Additional peaks are seen at *g* 8.2 and 5.6, along with a small but reproducible feature at *g* 9.6 and another small signal at *g* 2.0 which is likely a radical impurity. There are no signals that can be attributed to a low-spin ($S = 1/2$) iron(III) species, such as seen for the unmethylated analogue $[\text{Fe}^{\text{III}}([15]\text{aneN}_4)(\text{SPh})(\text{OOtBu})]^+$.

At first glance the intense signal at *g* 4.3 and the small feature at *g* 9.6 may be assigned to a purely rhombic high-spin Fe^{III} with $\lambda = E/D = 1/3$, while the features at *g* 8.2 and 5.6 could be associated with an intermediate λ value of ~ 0.1 , suggesting the presence of two different iron species. However, a recent, detailed EPR study by Nilges and Wilker on high-spin Fe^{III} -catecholate complexes shows that simulations of X-band EPR spectra based on only one high-spin Fe^{III} component with

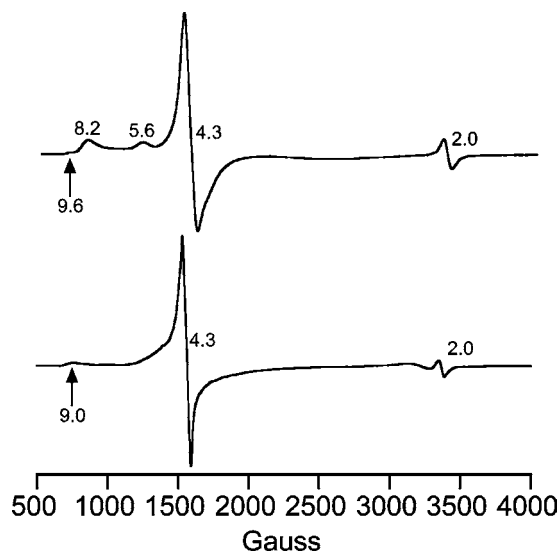


Figure 5. X-band EPR spectra of $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{SPh})(\text{OOtBu})]^+$ (top) and $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{OTf})(\text{OOtBu})]^+$ (bottom) in CH_2Cl_2 at 15 K. Experimental conditions (top, bottom): Frequency, 9.475 GHz; microwave power, 2 mW; modulation frequency, 100 kHz; modulation amplitude, 10.0 G; receiver gain, 5.02×10^3 .

$\lambda < 1/3$ reproduce all of the same spectral features seen for **2a**.⁶⁹ The EPR spectrum for **2a** was quantitated by double integration of the high-spin iron(III) region (600–2400 G) and comparison with a calibration curve constructed from an Fe^{III} -EDTA standard for an $S = 5/2$ ion.^{70,71} This analysis gives an 80% yield for **2a**, assuming the high-spin Fe^{III} signals arise only from **2a**. To further explore if the high-spin Fe^{III} EPR signals were coming from one species (i.e., **2a**) or multiple components, EPR spectra were monitored at different reaction times (30–120 min). As seen in Figure S2, the *g* 4.3, 5.6, and 8.2 peaks appear to broaden and shift over time. At the end of the time course a new feature at *g* ~ 6.7 is apparent, together with broad peaks at *g* ~ 5.6 and ~ 4.3 . Although a detailed analysis of these spectral changes is complicated due to the overlapping and broad nature of the features in this region, the peaks at *g* 8.2, 5.6, and 4.3 do not appear to interconvert or decay independently of each other. Double integrations at each time point give a relatively constant value of spins versus total iron content (79–87%). Thus these data are consistent with our assignment of the *g* 8.2, 5.6, and 4.3 peaks coming from a single species, although multiple species cannot be definitively ruled out. Taken together, these data are consistent with our assignment of **2a** as a high-spin alkylperoxo-iron(III) complex, $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{SPh})(\text{OOtBu})]^+$.

The spectrum for the triflate-ligated **3a** (Figure 5, bottom) is characterized by peaks at *g* 9.0 and 4.3, where the *g* 4.3 resonance extends out ~ 500 G to the high-field side, and can be assigned to a high-spin iron(III) complex following Nilges and Wilker.⁶⁹ The small signal at *g* 2.0 is likely a radical impurity as seen in the spectrum for **2a**. The EPR spectrum for **3a** is markedly different from that of **2a**, and in particular the dominant feature (*g* ~ 4.3) appears significantly broadened for **2a**. Qualitatively, this difference results from the triflate-coordinated complex more closely approaching ideal rhombic

(69) Weisser, J. T.; Nilges, M. J.; Sever, M. J.; Wilker, J. J. *Inorg. Chem.* **2006**, *45*, 7736–7747.

(70) Aasa, R.; Vänngård, T. *J. Magn. Reson.* **1975**, *19*, 308–315.

(71) Bou-Abdallah, F.; Chasteen, N. D. *J. Biol. Inorg. Chem.* **2008**, *13*, 15–24.

Table 3. Vibrational, UV–Vis, and EPR Data for High-Spin Fe^{III}–OOR Complexes

high-spin Fe ^{III} –OOR complexes ^a	$\nu(\text{Fe–O})$ (cm ⁻¹)	$\nu(\text{O–O})$ (cm ⁻¹)	λ (nm)	EPR <i>g</i> values	ref
[Fe ^{III} (Me ₄ [15]aneN ₄)(SPh)(OO <i>t</i> Bu)] ⁺ (2a)	584	843, 872	650	9.6, 8.2, 5.6, 4.3	<i>b</i>
[Fe ^{III} (Me ₄ [15]aneN ₄)(OTf)(OO <i>t</i> Bu)] ⁺ (3a)	612	871	600	9.0, 4.3	<i>b</i>
[Fe ^{III} (6Me ₃ TPA)(OO <i>t</i> Bu)(OH)] ⁺	637	842, 877	562	4.3	76
[Fe ^{III} (6Me ₃ TPA)(OO <i>t</i> Bu)(H ₂ O)] ²⁺	612	829, 867	562	4.3	76
[Fe ^{III} (6Me ₂ TPA)(OO <i>t</i> Bu)(H ₂ O)] ²⁺	647	843, 881	552	4.3	76
[Fe ^{III} (6MeTPA)(OO <i>t</i> Bu)(H ₂ O)] ²⁺	682	790	600	4.3, 2.2, 2.12, 1.97 ^c	76
[Fe ^{III} (L ₈ Py ₂)(OO <i>t</i> Bu)(OTf)] ⁺	627	833, 870	580	8.8, 5.2, 3.2	20
[Fe ^{III} (L ₈ Py ₂)(OO <i>t</i> Bu)(OBz)] ⁺	623	832, 873	545	---	20
[Fe ^{III} (L ₈ Py ₂)(OO <i>t</i> Bu)(SAr)] ⁺	623	830, 874	510	---	20
[Fe ^{III} (L ₈ Py ₂)(OO <i>t</i> Bu)(OPy)] ⁺	623	831, 878	550	---	20
[Fe ^{III} (bppa)(OO <i>t</i> Bu)] ²⁺	629	873, 838	613	7.58, 5.81, 4.25, 1.82	77
[Fe ^{III} (bppa)(OOCm)] ²⁺	639	878, 838	585	7.76, 5.65, 4.20, 1.78	77

^a Abbreviations: TPA = tris(2-pyridylmethyl)amine; L₈Py₂ = *N,N*-bis(2-pyridylmethyl)-1,5-diazacyclooctane; bppa = bis(6-pivalamido-2-pyridylmethyl)(2-pyridylmethyl)amine. ^b This work. ^c Mixed high-spin and low-spin species.

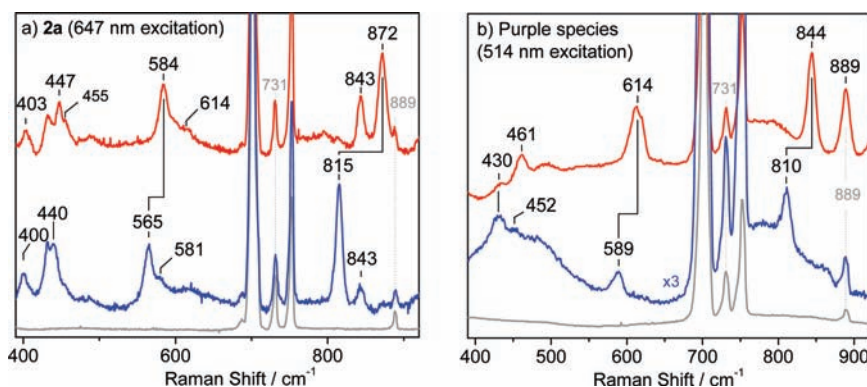


Figure 6. RR spectra of (a) [Fe^{III}(Me₄[15]aneN₄)(SPh)(OO*t*Bu)]⁺ (**2a**) generated from *t*Bu¹⁶O¹⁶OH (top, red) and *t*Bu¹⁸O¹⁸OH (middle, blue), and [Fe^{III}(Me₄[15]aneN₄)(SPh)(BPh₄)]⁺ (**2**) (bottom, gray) obtained with 647-nm laser excitation and (b) the purple species observed following the decay of **2a** generated from *t*Bu¹⁶O¹⁶OH (top, red) and *t*Bu¹⁸O¹⁸OH (middle, blue) and **2** (bottom, gray) obtained with 514-nm laser excitation. Spectra recorded on frozen solutions (CH₂Cl₂) at ~110 K. In each graph the spectra are normalized on Raman signals from CH₂Cl₂.

symmetry, while the thiolate-coordinated complex is more distorted. We choose not to speculate as to the electronic or geometrical origins of this effect but simply note that the difference in axial ligands (thiolate vs triflate) has a distinct effect on the electronic structure of the Fe^{III}–OOR unit as reflected in the EPR spectra.

The major finding to emerge from the EPR data is that *N*-methylation of [15]aneN₄ leads to a high-spin ground state (*S* = 5/2) for the iron(III)-alkylperoxo species. Previously, it was shown by EPR that the nonmethylated complexes [Fe^{III}([15]aneN₄)(SAr)(OOR)]⁺ were all low-spin Fe^{III} (*S* = 1/2). This result is consistent with our findings regarding the influence of the methyl groups; methylation causes a significant weakening of the amine donor strength in [15]aneN₄, and thus alkylation at nitrogen provides access to high-spin Fe^{III}–OOR complexes.

Resonance Raman Spectroscopy. The RR spectra of **2a** obtained at 110 K are shown in Figure 6. With a 647-nm laser excitation that coincides with the absorption maximum, the RR spectrum exhibits strong resonance-enhanced vibrations near 450, 584, 843, and 872 cm⁻¹. Preparing **2a** with *t*Bu¹⁸O¹⁸OH affects all of these RR signals. The bands near 450 cm⁻¹ are only moderately perturbed by ¹⁸O-labeling and are assigned to (C–C–C) and (C–C–O) deformation modes of the alkylperoxo ligand as in previous RR characterizations of iron(III)-alkylperoxo species.^{15,16,25,34} The 584-cm⁻¹ band which shifts –19 cm⁻¹ with *t*Bu¹⁸O¹⁸OH is assigned to the $\nu(\text{Fe–O})$ (Table 3). Finally, the two strong bands at 872 and 843 cm⁻¹ are replaced after labeling with *t*Bu¹⁸O¹⁸OH by a weak 843-cm⁻¹ band and a strong band at 815 cm⁻¹ readily assigned to the $\nu(^{18}\text{O–}^{18}\text{O})$

mode. The enhancement of the 843 cm⁻¹ band in unlabeled **2a** most likely originates from Fermi coupling between $\nu(\text{O–O})$ and $\nu(\text{C–C})$ of the OO*t*Bu group as previously noted for related complexes.^{15,72} In agreement with this analysis, using a $\nu(^{18}\text{O–}^{18}\text{O}) = 815$ cm⁻¹ to calculate the $\nu(^{16}\text{O–}^{16}\text{O})$ assuming a simple diatomic oscillator leads to a 864.5 cm⁻¹ value, intermediate between the 843 and 872 cm⁻¹ bands observed in **2a**. These $\nu(\text{O–O})$ values are comparable to other frequencies reported for high-spin alkylperoxo complexes (Table 3), but the $\nu(\text{Fe–O})$ of **2a** is 40 to 60 cm⁻¹ lower than those observed for other high-spin alkylperoxo complexes. This $\nu(\text{Fe–O})$ is also significantly lower than that seen for the low-spin complex [Fe^{III}([15]aneN₄)(SPh)(OO*t*Bu)]⁺ ($\nu(\text{Fe–O}) = 615$ cm⁻¹).²⁵ As previously seen with the unmethylated low-spin Fe^{III}–OOR complexes, the RR spectra of **2a** show no evidence of Fe–S stretching vibrations in the 300 to 400 cm⁻¹ region (Figure S3).

The purple species produced by the decay of **2a** contributes only weakly to RR spectra obtained with 647-nm excitation (see Supporting Information), but the RR spectra obtained with 514-nm excitation for samples where the purple species was intentionally maximized show strong $\nu(\text{Fe–O})$ and $\nu(\text{O–O})$ vibrations at 614 and 844/889 cm⁻¹, respectively (Figure 6b). At this excitation wavelength, the RR modes of **2a** are only weakly enhanced (Figure S4). As with most iron(III)-alkylperoxo species, the RR spectrum of the purple species shows evidence of Fermi coupling between $\nu(\text{O–O})$ and, presumably,

(72) Lehnert, N.; Fujisawa, K.; Solomon, E. I. *Inorg. Chem.* **2003**, *42*, 469–481.

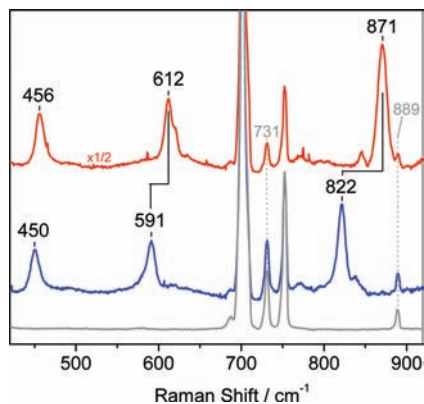


Figure 7. RR spectra of $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{OTf})(\text{OOtBu})]^+$ (**3a**) generated from $t\text{Bu}^{16}\text{O}^{16}\text{OH}$ (top, red) and $t\text{Bu}^{18}\text{O}^{18}\text{OH}$ (middle, blue), and $[\text{Fe}^{\text{II}}(\text{Me}_4[15]\text{aneN}_4)(\text{OTf})_2]$ (**3**) (bottom, gray). Spectra were recorded on frozen solutions (CH_2Cl_2) at ~ 110 K with 647-nm laser excitation.

the $\nu(\text{C}-\text{C})$ of the OOtBu group. Labeling of the alkylperoxy moiety with ^{18}O shifts the $\nu(\text{O}-\text{O})$ mode to 810 cm^{-1} , out of coupling range with the nonresonant $\nu(\text{C}-\text{C})$ mode. Comparing $\nu(^{18}\text{O}-^{18}\text{O})$ and $\nu(\text{Fe}-\text{O})$ in the purple species and **2a** reveals a modest downshift of the $\text{O}-\text{O}$ stretch (-5 cm^{-1}) and a $+30\text{ cm}^{-1}$ upshift of the $\nu(\text{Fe}-\text{O})$ in the purple species which is in line with the loss of a strong thiolate *trans* effect. The nature of the sixth ligand in this species has not been identified.⁷³

The RR spectra for the blue species **3a**, prepared from the reaction of $[\text{Fe}^{\text{II}}(\text{Me}_4[15]\text{aneN}_4)(\text{OTf})_2]$ (**3**) and $t\text{BuOOH}$ at $-23\text{ }^\circ\text{C}$, are shown in Figure 7. The RR spectra, obtained with a 647-nm excitation, show a set of $\nu(\text{Fe}-\text{O})$ and $\nu(\text{O}-\text{O})$ vibrations at 612 and 871 cm^{-1} that downshift to 591 and 822 cm^{-1} with $t\text{Bu}^{18}\text{O}^{18}\text{OH}$, respectively. These data confirm the assignment of **3a** as the iron(III)-alkylperoxy complex $[\text{Fe}^{\text{III}}(\text{Me}_4[15]\text{aneN}_4)(\text{OTf})(\text{OOtBu})]^+$. In contrast with the RR spectra of **2a**, the region corresponding to the $(\text{C}-\text{C}-\text{C})/(\text{C}-\text{C}-\text{O})$ deformation modes of **3a** shows a single band at 456 cm^{-1} that shifts -6 cm^{-1} with $t\text{Bu}^{18}\text{O}^{18}\text{OH}$. Importantly, the lack of a *trans* thiolate ligand in **3a** results in a $\nu(\text{Fe}-\text{O})$ 18 cm^{-1} higher than in **2a**, while its $\nu(^{18}\text{O}-^{18}\text{O})$ mode upshifts 7 cm^{-1} compared to **2a** (Table 3).

As pointed out in the Introduction, the spin states of $\text{Fe}^{\text{III}}-\text{OOR}(\text{H})$ complexes have been suggested to strongly influence the $\text{Fe}-\text{O}$ and $\text{O}-\text{O}$ vibrations and accompanying bond strengths. Specifically, a low-spin ground state ($S = 1/2$) should result in relatively strong $\text{Fe}-\text{O}$ and weak $\text{O}-\text{O}$ bonds, while a high-spin ground state ($S = 5/2$) should lead to weak $\text{Fe}-\text{O}$ and strong $\text{O}-\text{O}$ bonds.^{15,16,25} The low-spin complexes $[\text{Fe}^{\text{III}}([15]\text{aneN}_4)(\text{SAr})(\text{OOR})]^+$ exhibited low $\nu(\text{Fe}-\text{O})$ s between 600 and 623 cm^{-1} that did not follow the expected trend for low-spin vs high-spin $\text{Fe}^{\text{III}}-\text{OOR}$ complexes. These unusually low $\nu(\text{Fe}-\text{O})$ s were assigned to a *trans* effect of the thiolate donor, and it was shown that the electron-donating power of the different thiolate ligands correlated closely with the $\text{Fe}-\text{O}$ vibrational frequencies. The ^{18}O -downshifts of the $\nu(\text{Fe}-\text{O})$ and $\nu(\text{O}-\text{O})$ bands for the low spin, unmethylated complexes matched those predicted for simple diatomic oscillators and

permitted a direct interpretation of vibrational energies in terms of bond strengths. For the high-spin species **2a**, the observed isotopic downshifts deviate from calculated values for simple diatomic models ($\Delta\nu(\text{Fe}-^{16/18}\text{O})\text{ exptl (calc)} = -19 (-28)\text{ cm}^{-1}$ and Fermi coupled $\nu(^{16}\text{O}-^{16}\text{O})$ doublet), and therefore interpreting vibrational frequency in terms of bond strengths is less straightforward. However, the present work demonstrates that both the tetramethylated and nonmethylated thiolate-ligated SOR model complexes do follow the expected trend in $\nu(\text{Fe}-\text{O})$ and $\nu(\text{O}-\text{O})$ vs iron spin state, with the important finding that $\nu(\text{Fe}-\text{O})$ is dramatically weakened by the axial thiolate donor for both high-spin and low-spin species. In contrast, the $\nu(\text{O}-\text{O})$ frequencies appear to be only marginally affected by the thiolate ligand. To our knowledge this study is the first in which the influence of a ligand *trans* to a peroxo moiety has been evaluated for a structurally equivalent pair of high-spin/low-spin peroxo-iron(III) complexes.

The triflate-ligated complex **3a** and the thiolate-ligated complex **2a** exhibit comparable $\nu(\text{O}-\text{O})$ s (as judged by the $\nu(^{18}\text{O}-^{18}\text{O})$ modes which show no evidence of Fermi coupling) but different $\nu(\text{Fe}-\text{O})$ s. The more weakly donating triflate ligand leads to an $\sim 30\text{ cm}^{-1}$ increase in $\nu(\text{Fe}-\text{O})$ compared to the thiolate ligand. These results contrast with what was seen in the high-spin complexes $[\text{Fe}^{\text{III}}(\text{L}_8\text{Py}_2)(\text{OOtBu})(\text{X})]^+$ ($\text{X} = \text{OTf}, \text{OBz}, \text{SAr}, \text{OPy}$), where the iron-alkylperoxy $\nu(\text{Fe}-\text{O})$ is largely insensitive to a change in the nature of the *trans* ligand (Table 3). For these complexes, only the $\nu(\text{Fe}-\text{O})$ of the triflate-ligated species exhibited a small upshift of 4 cm^{-1} compared to the other complexes. The lack of sensitivity of $\nu(\text{Fe}-\text{O})$ to the nature of the sixth ligand was not discussed.²⁰

Our results on the high-spin complexes parallel our findings for the low-spin complexes, in which $\nu(\text{Fe}-\text{O})$ modes are sensitive to the identity of the *trans* donor ligand, while $\nu(\text{O}-\text{O})$ modes are only marginally affected. These results support the strikingly similar findings that have been obtained for (hydro)-peroxo-iron(III) adducts of SOR, where a weakening of the $\text{S}^{\text{Cys}}-\text{Fe}^{\text{III}}$ bond correlates with a strengthening of the $\text{Fe}-\text{O}$ bond but has little or no effect on the $\text{O}-\text{O}$ bond.^{6,23} In fact, there is a growing body of evidence to suggest that the nature of the $\text{S}-\text{Fe}$ interaction in SOR plays a critical role in catalysis.^{3,23,74,75} Our findings and those on the enzyme are in agreement and show that the traditional “push effect” invoked for heme enzymes, where the thiolate donor helps to weaken the $\text{O}-\text{O}$ bond and thereby favor $\text{O}-\text{O}$ bond cleavage, is not observed in either our low-spin or high-spin nonheme iron models, or in the native enzyme itself. However, these findings do not rule out a possible push effect on excited state species (e.g., transition state in $\text{O}-\text{O}$ bond cleavage) during the formation and decay of an iron-peroxide.

Summary and Conclusions

The alkylated ligand $\text{Me}_4[15]\text{aneN}_4$ has been used to prepare a new thiolate-ligated, $(\text{N}_4\text{S})\text{Fe}^{\text{II}}$ complex as an accurate structural model of the reduced form of SOR. This complex adds to the small but growing number of thiolate-ligated SOR

(73) Experiments aimed at better characterizing the status of the iron(III) coordination *trans* to the alkylperoxy ligand in the purple species were inconclusive. Specifically, addition of sodium benzenesulfinate, a likely oxidation product of the phenylthiolate ligand, to **3a** did not convert this species to the purple complex (514 nm). Similarly, addition of sodium triflate to the purple species did not convert the latter species to **3a**.

(74) Clay, M. D.; Cosper, C. A.; Jenney, F. E.; Adams, M. W. W.; Johnson, M. K. *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 3796–3801.

(75) Todorovic, S.; Rodrigues, J. V.; Pinto, A. F.; Thomsen, C.; Hildebrandt, P.; Teixeira, M.; Murgida, D. H. *Phys. Chem. Chem. Phys.* **2009**, *11*, 1809–1815.

(76) Zang, Y.; Kim, J.; Dong, Y. H.; Wilkinson, E. C.; Appelman, E. H.; Que, L., Jr. *J. Am. Chem. Soc.* **1997**, *119*, 4197–4205.

(77) Wada, A.; Ogo, S.; Watanabe, Y.; Mukai, M.; Kitagawa, T.; Jitsukawa, K.; Masuda, H.; Einaga, H. *Inorg. Chem.* **1999**, *38*, 3592–3593.

models. Previously we prepared a series of structurally analogous SOR models with the nonalkylated ligand [15]aneN₄. We found that the addition of methyl groups on the amine donors has a dramatic impact on the donating properties of the macrocycle, making Me₄[15]aneN₄ a significantly weaker donor than [15]aneN₄. We have shown that the tetramethylated iron(II) complex is capable of producing the metastable alkylperoxo species [Fe^{III}(Me₄[15]aneN₄)(SPh)(OO*t*Bu)]⁺ (**2a**), despite the potential steric crowding of the metal center from the methyl groups. This complex is a rare example of an iron-peroxo species that contains a thiolate donor in the coordination sphere. Importantly, alkylation at nitrogen induces a spin state change for the alkylperoxo species from low-spin to high-spin iron(III). The change in spin state causes a dramatic shift in the Fe–O and O–O vibrations of the Fe–OO*t*Bu unit, in which $\nu(\text{Fe–O})$ drops to lower energy by $\sim 30\text{ cm}^{-1}$ and $\nu(\text{O–O})$ increases by $\sim 60\text{ cm}^{-1}$. The $\nu(\text{Fe–O})$ for this high-spin alkylperoxo species **2a** is lower than that for all other known high-spin alkylperoxo complexes lacking a thiolate ligand.

This work has allowed for a direct comparison of iron-peroxo species with low-spin and high-spin iron(III) in nearly identical ligand environments. Previously, our data for the thiolate-ligated low-spin Fe^{III}–OOR model complexes, and in particular their very low $\nu(\text{Fe–O})$, suggested that these models may not follow the trend of other low-spin Fe^{III}–OOR species that yield strong Fe–O and weak O–O bonds compared to high-spin homologues. However, the characterization of the thiolate-ligated high-spin Fe^{III}–OOR **2a** shows an equally low $\nu(\text{Fe–O})$ when compared to other high-spin Fe–OOR species. Thus, these studies demonstrate that the addition of the *trans* thiolate ligand

in both low-spin and high-spin species induces a large weakening of the $\nu(\text{Fe–O})$ without a major change in $\nu(\text{O–O})$. A new nonthiolate-ligated alkylperoxo complex was also generated, [Fe^{III}(Me₄[15]aneN₄)(OTf)(OO*t*Bu)]⁺ (**3a**), and exhibited an $\sim 30\text{-cm}^{-1}$ upshift in $\nu(\text{Fe–O})$ but only a 7-cm^{-1} upshift in $\nu(\text{O–O})$. These data provide strong evidence that the *trans* donor ligand in Fe^{III}–OOR species has an important role in fine-tuning $\nu(\text{Fe–O})$. Taken together, our results indicate that both *spin state* and *trans-thiolate ligation* play a critical role in determining the nature of the peroxo–iron interaction in models and, by analogy, in SOR. The His₄Cys iron-binding site in SOR, as opposed to the porphyrinate/Cys ligand set found in cytochrome P450, may have been selected to ensure the formation of a high-spin Fe^{III}–OO(H) intermediate with a relatively weak Fe–O bond, thereby favoring the release of H₂O₂ over O–O bond cleavage.

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Supporting Information Available: Additional RR, UV–vis and EPR spectra, and X-ray structure file for complex **2** (CIF). This material is available free of charge via the Internet at <http://pubs.acs.org>

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